

# Literature Report

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**Reporter: 许宁**  
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# Distinct fission signatures predict mitochondrial degradation or biogenesis

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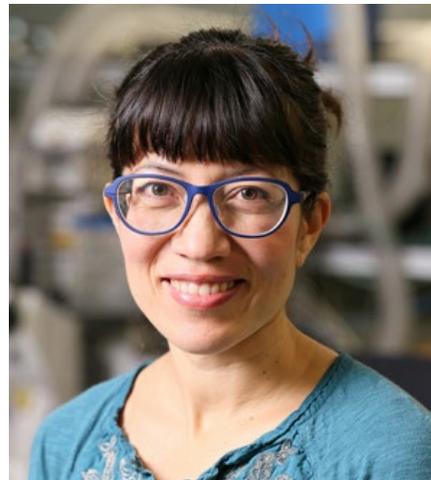
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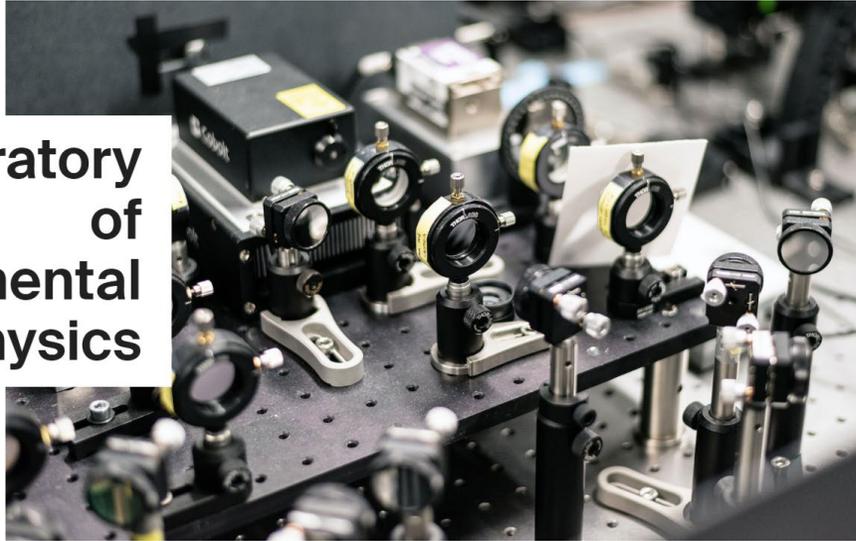


Postdoctoral Researcher  
Laboratory of Experimental Biophysics



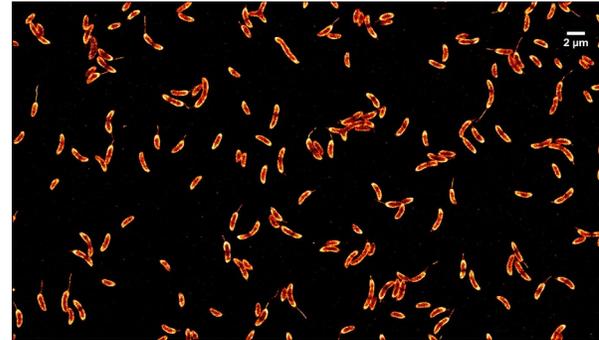
1997 B.A. Physics and Mathematics, Rice University  
2004 PhD Physics, Harvard University.  
2004-2006 Postdoctoral fellow, Department of Chemical  
Engineering, MIT, Cambridge, MA  
2006-2009 National Research Council postdoctoral fellow, NICHD,  
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2009-2016 Assistant professor, Department of Physics, EPFL,  
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# Laboratory of Experimental Biophysics



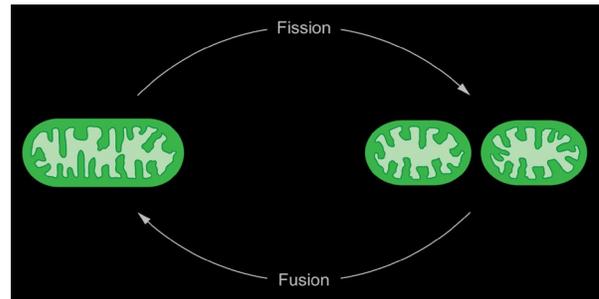
We develop and use automated super-resolution fluorescence imaging techniques combined with **live cell imaging** and **single molecule tracking** to determine how the dynamics of protein assembly are coordinated.

## Large field-of-view super-resolution microscopy

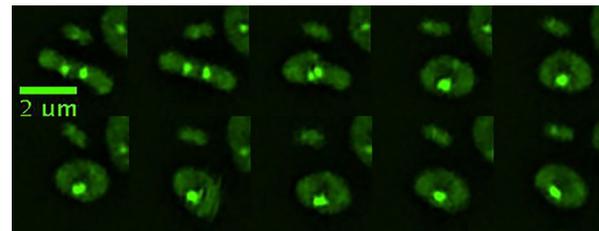


a large sample area ( $\sim 150 \times 150 \mu\text{m}^2$ )

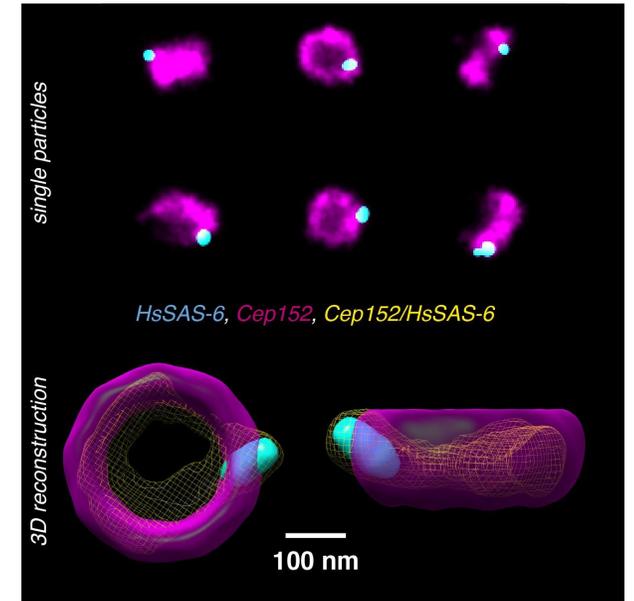
## Mitochondrial dynamics



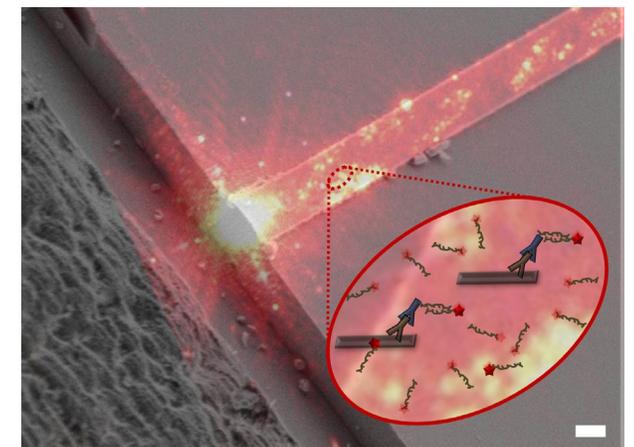
## Organization of mitochondrial gene expression



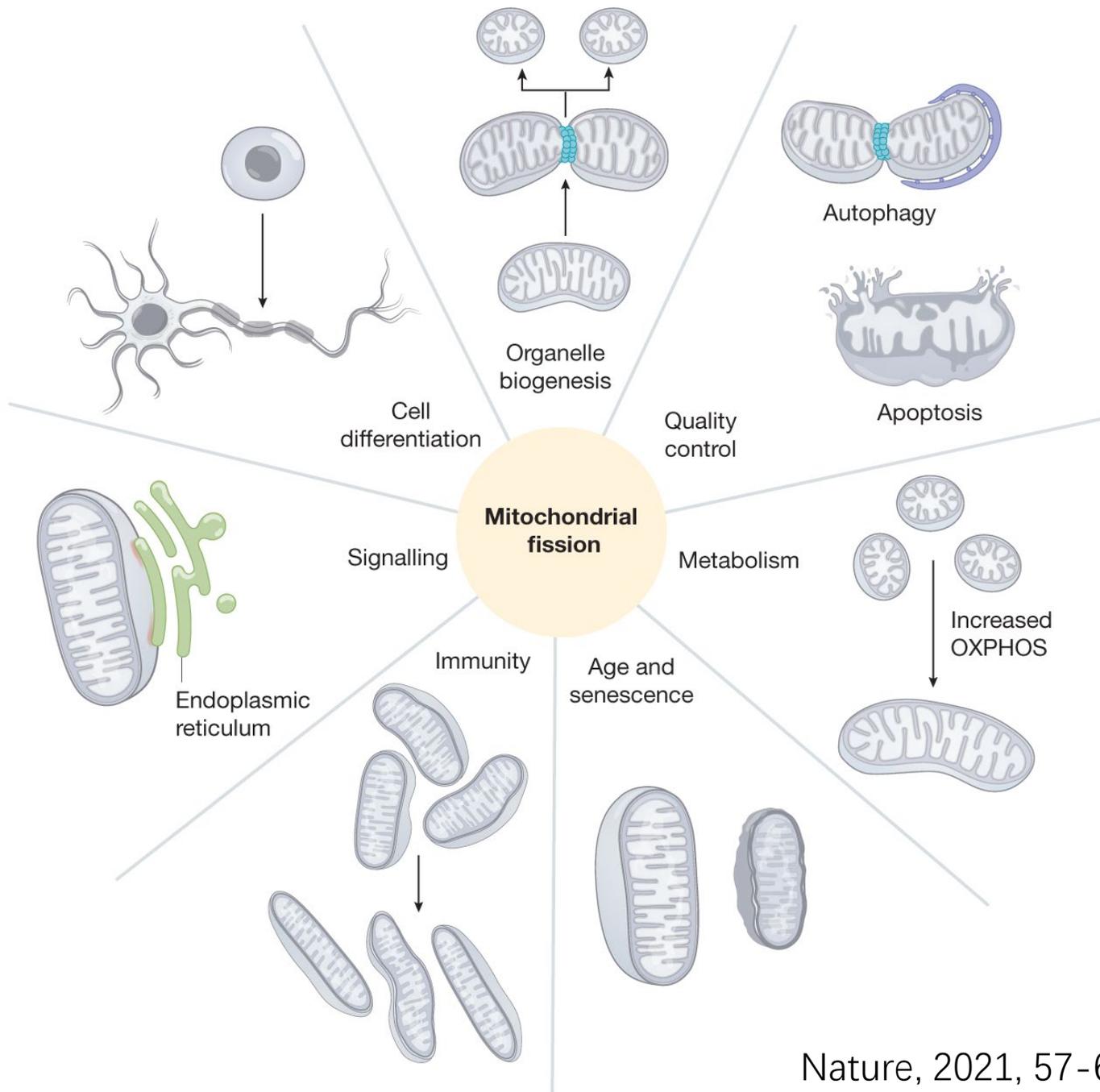
## Multicolor 3D single particle reconstruction



## Waveguide TIRF for high-throughput DNA-PAINT

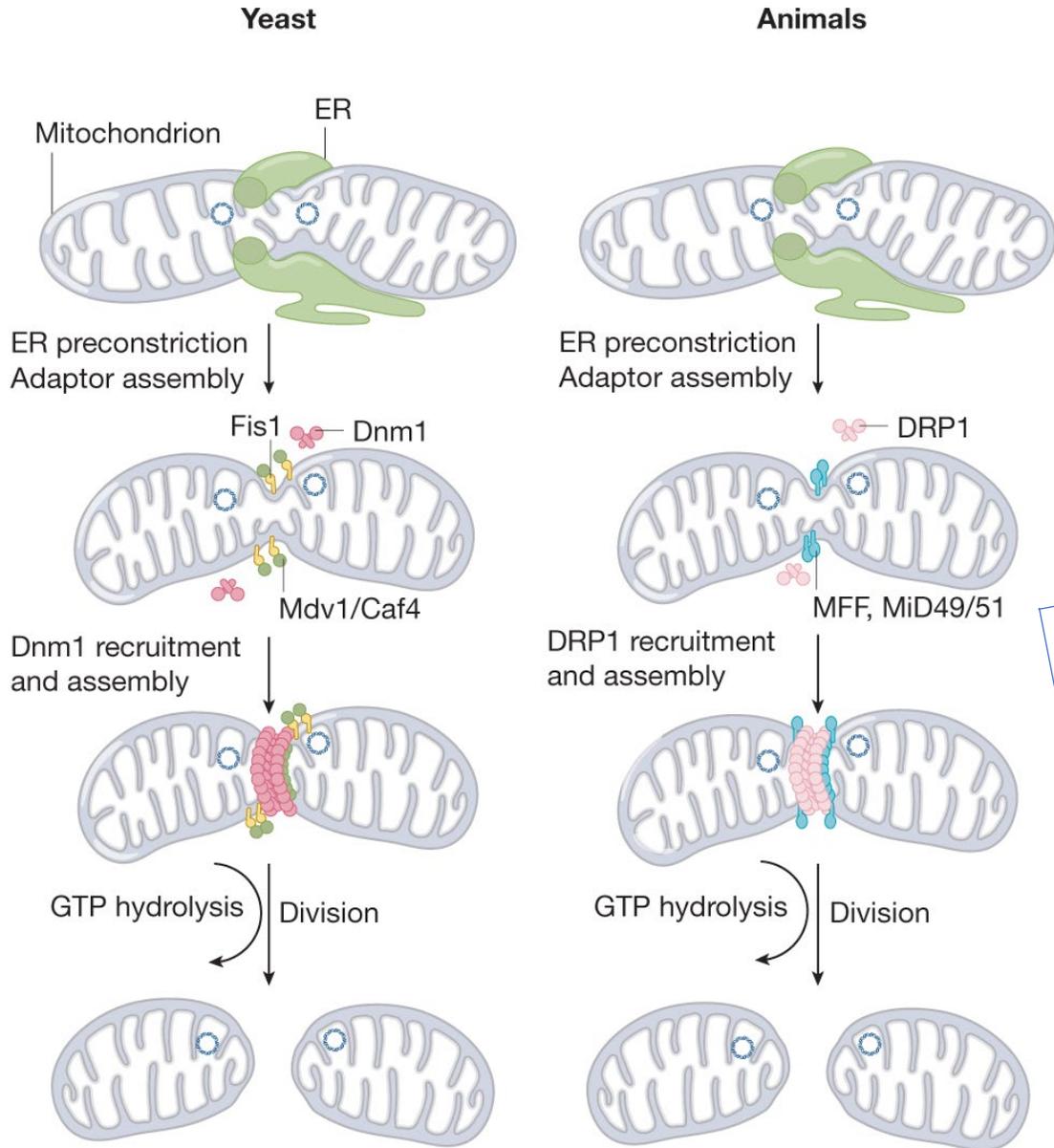


# Cellular and physiological importance of the mitochondrial divisome.

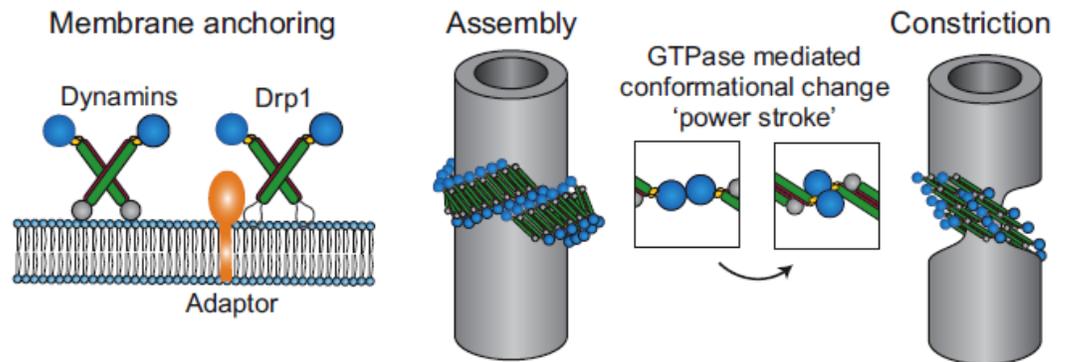
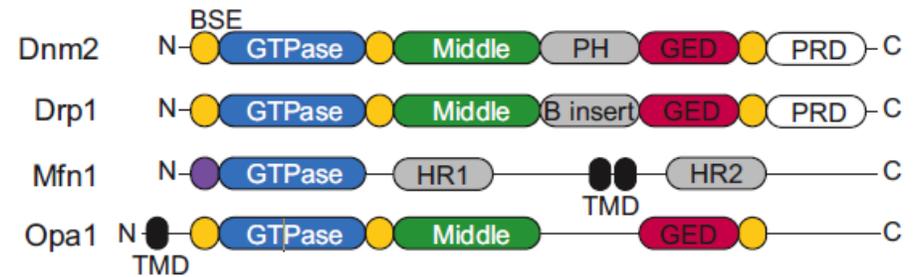


线粒体分裂失调:

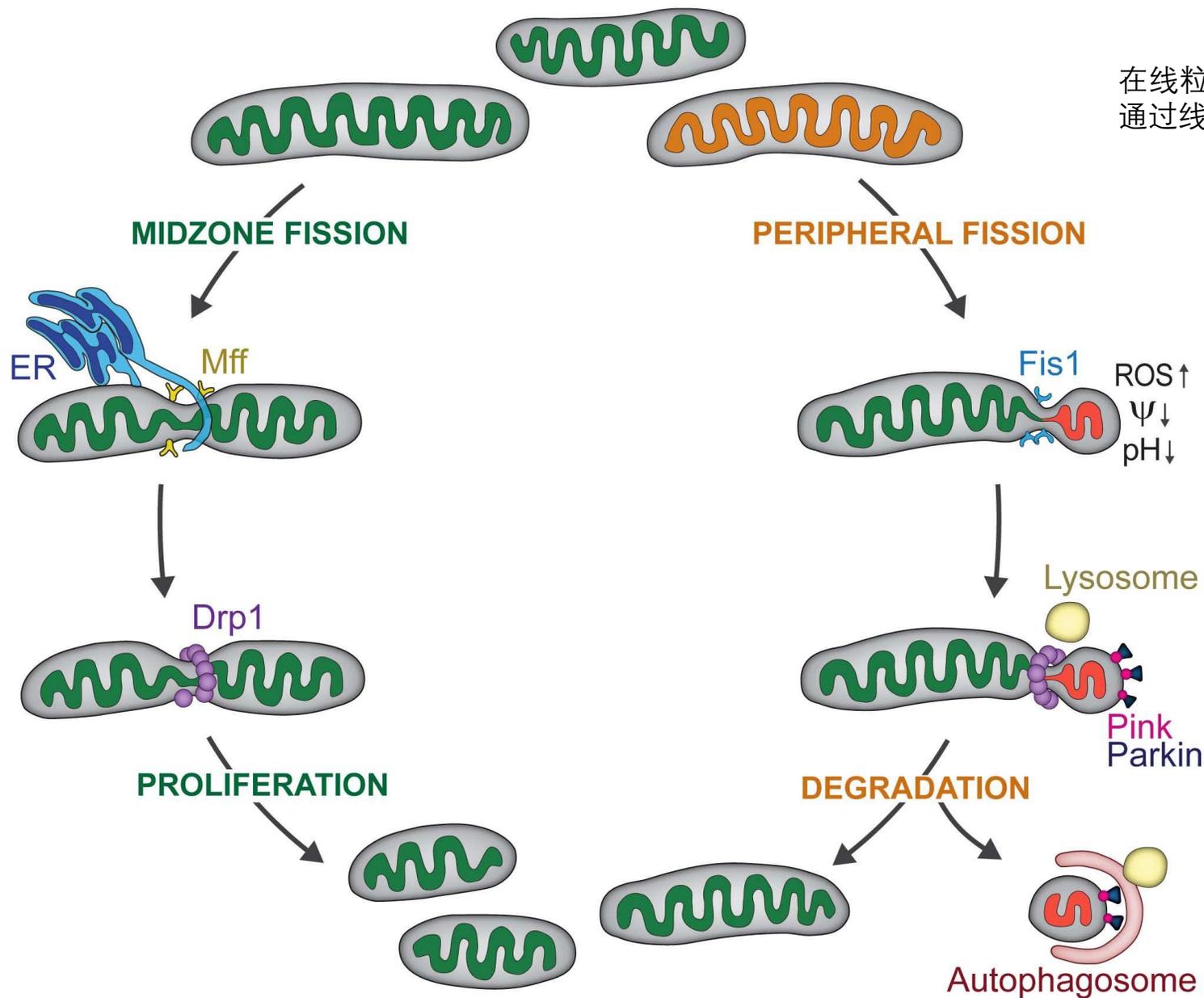
神经退行性疾病  
心血管疾病  
癌症



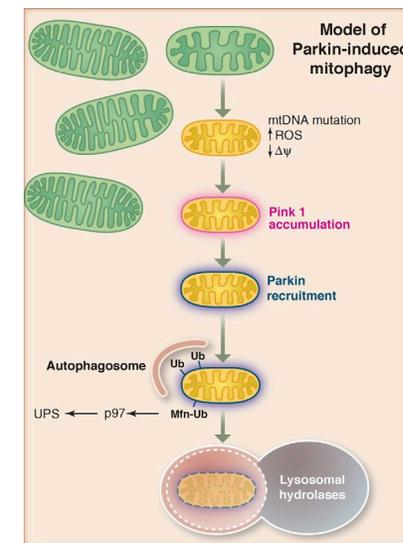
the GTPase of the dynamin superfamily of proteins  
动力蛋白相关蛋白1 (DRP1)

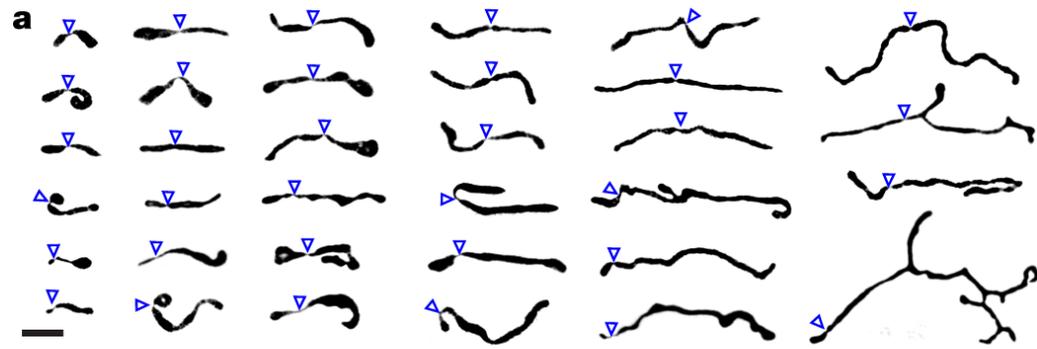


# Distinct fission signatures predict mitochondrial degradation or biogenesis



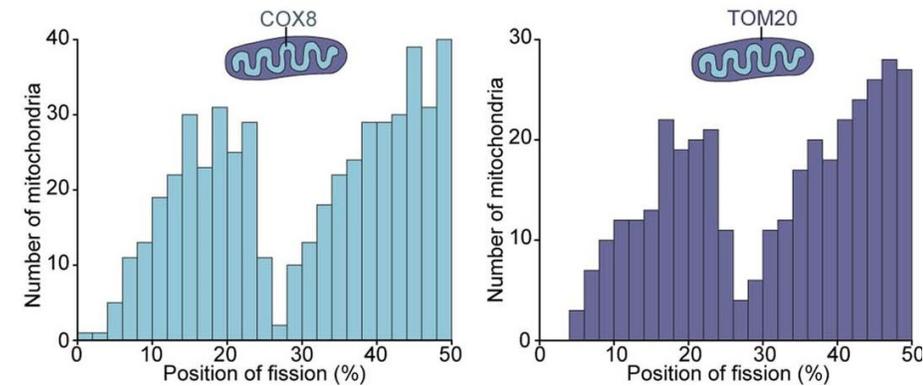
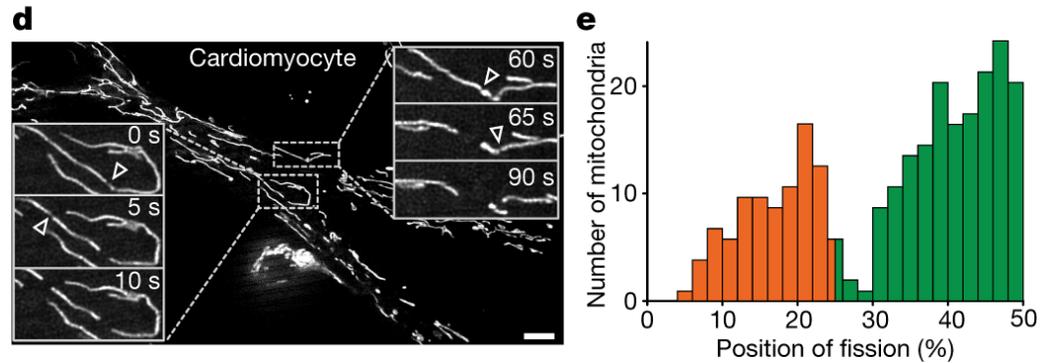
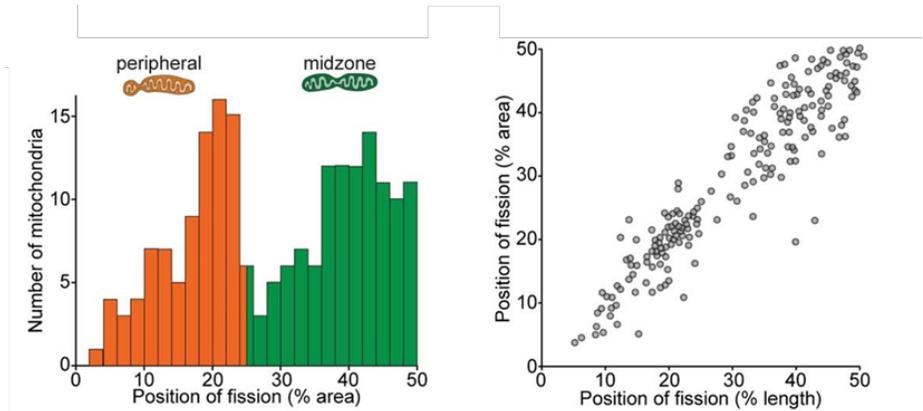
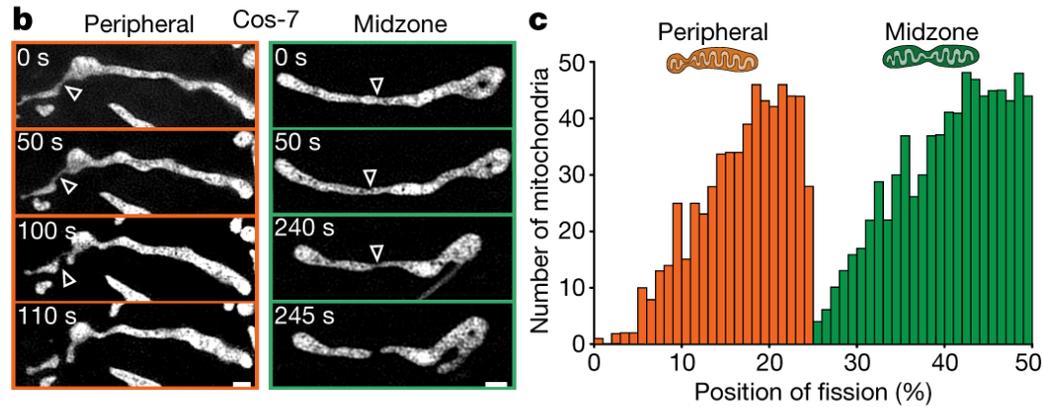
在线粒体的生命周期中，分裂既能使新的线粒体产生，又能通过线粒体自噬作用清除功能失调的线粒体。





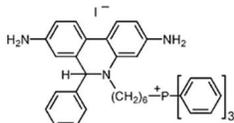
Peripheral divisions (less than 25% from a tip)

Midzone divisions (within the central 50%)

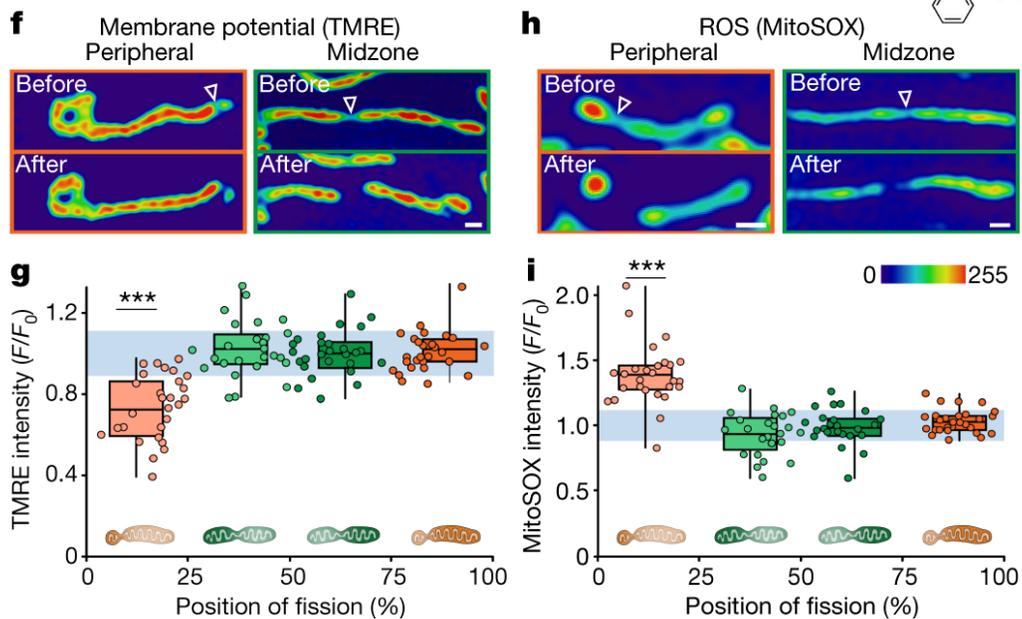


**Fig. 1: Mitochondrial fissions are bimodally positioned and linked to distinct physiologies.** **a**, Gallery of mitochondria one frame before division from structured illumination microscopy (SIM) movies, binarized. **b**, Time-lapse SIM sequence of a peripheral and a midzone fission of mitochondria (Mitotracker Green). **c**, Histogram of fission positions relative to the total mitochondrial length ( $n = 1,393$  fissions pooled from multiple datasets). Fissions occurring near the tip (orange, 0–25%) are termed ‘peripheral’; those near the centre (green, 25–50%) are termed ‘midzone’. **d**, Instant SIM (iSIM) of mitochondria (Mitotracker Green) in primary mouse cardiomyocytes. Insets: time-lapse sequences from indicated boxes. **e**, Histogram of the relative position of fission in cardiomyocyte mitochondria ( $n = 381$  fissions), as in **c**.

# Dysfunction precedes peripheral fission



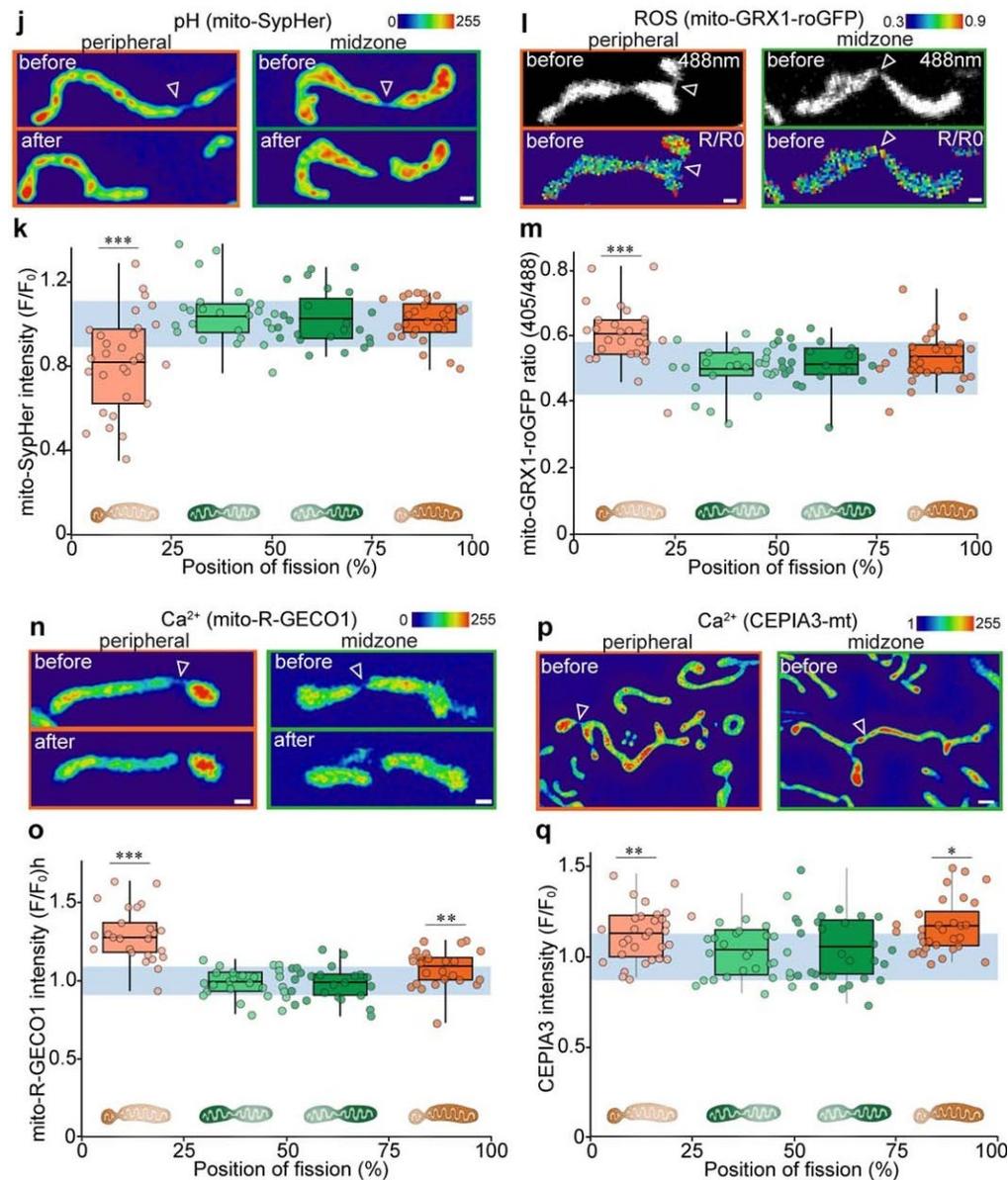
几何上不同的分裂类型是否反映了潜在的生理差异



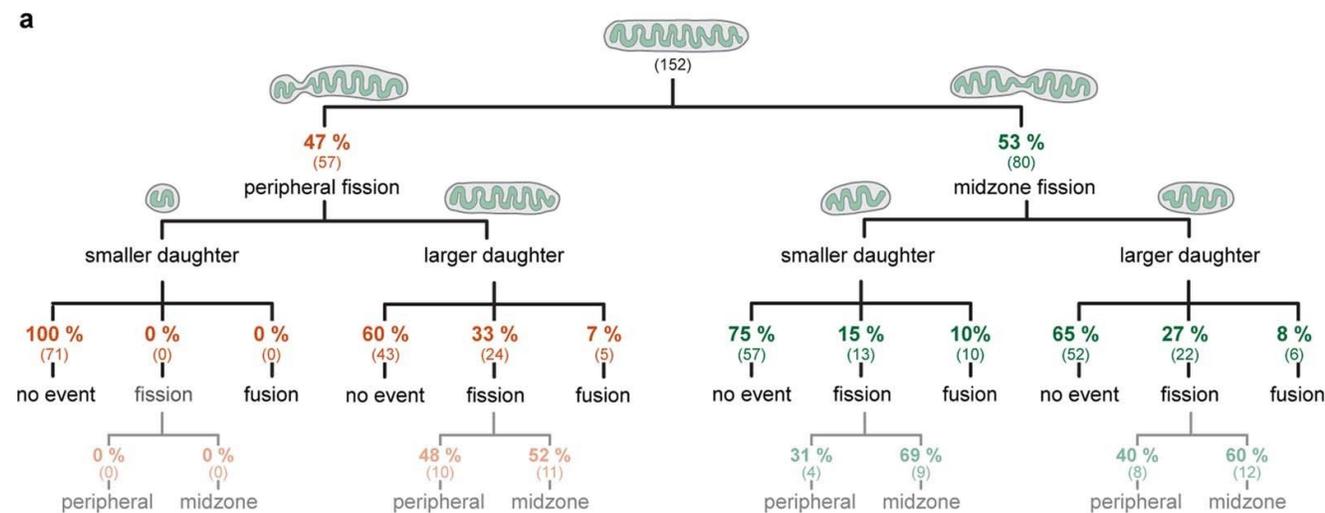
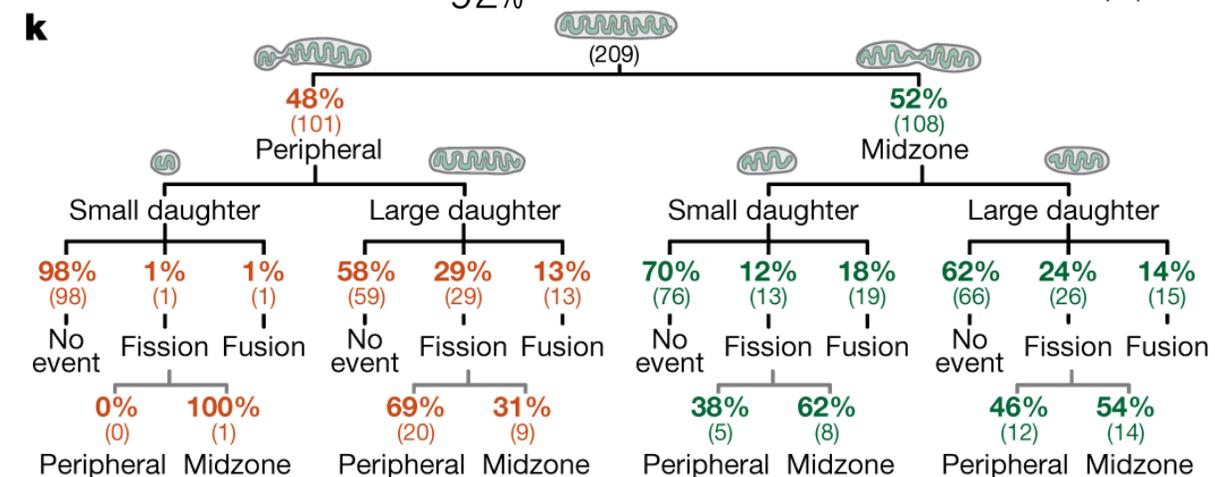
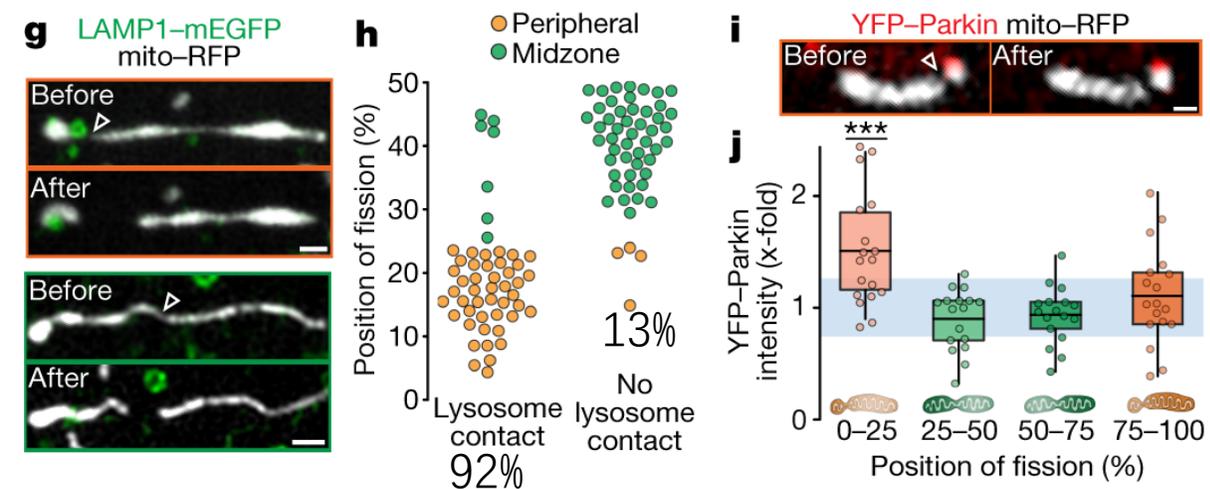
**Fig. 1: Mitochondrial fissions are bimodally positioned and linked to distinct physiologies.** **f**, Mitochondrial membrane potential before and after a peripheral or midzone fission from SIM movies of tetramethylrhodamine ethyl ester (TMRE)-stained mitochondria. **g**, Normalized TMRE intensity as a function of relative position of fission, measured immediately before fission ( $n = 56$  fissions). **h**, MitoSOX labelling reveals ROS levels before and after a peripheral or midzone fission. **i**, Normalized MitoSOX intensity as a function of relative fission position, measured immediately before fission ( $n = 52$  fissions).

Midzone divisions  
分裂前后的生理状态没有差异

Peripheral divisions  
分裂前:  $Ca^{2+}$ 和ROS浓度增加,膜电位和pH降低



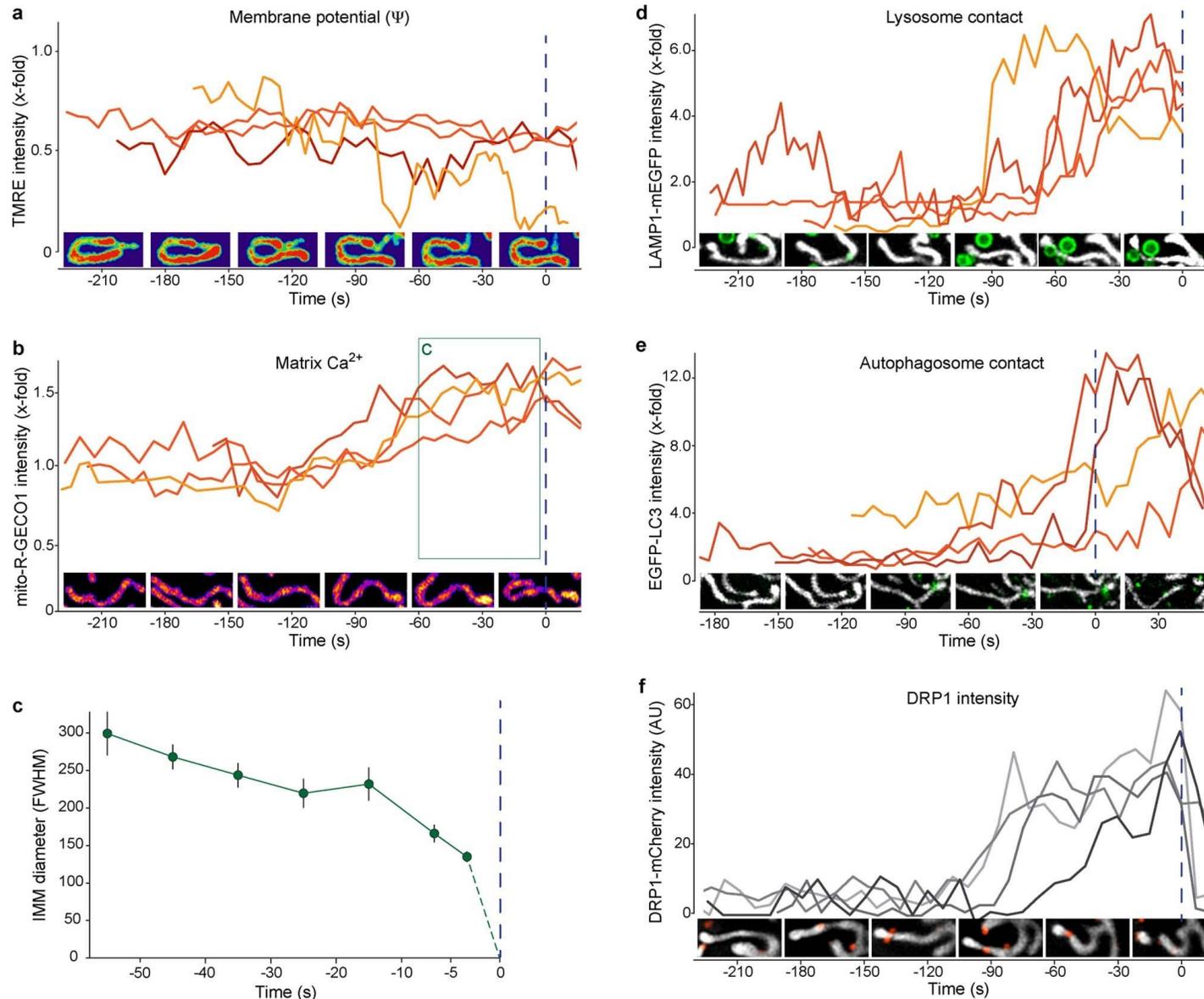




**Fig. 2: Midzone and peripheral fissions differ in mitochondrial DNA content and fates.**

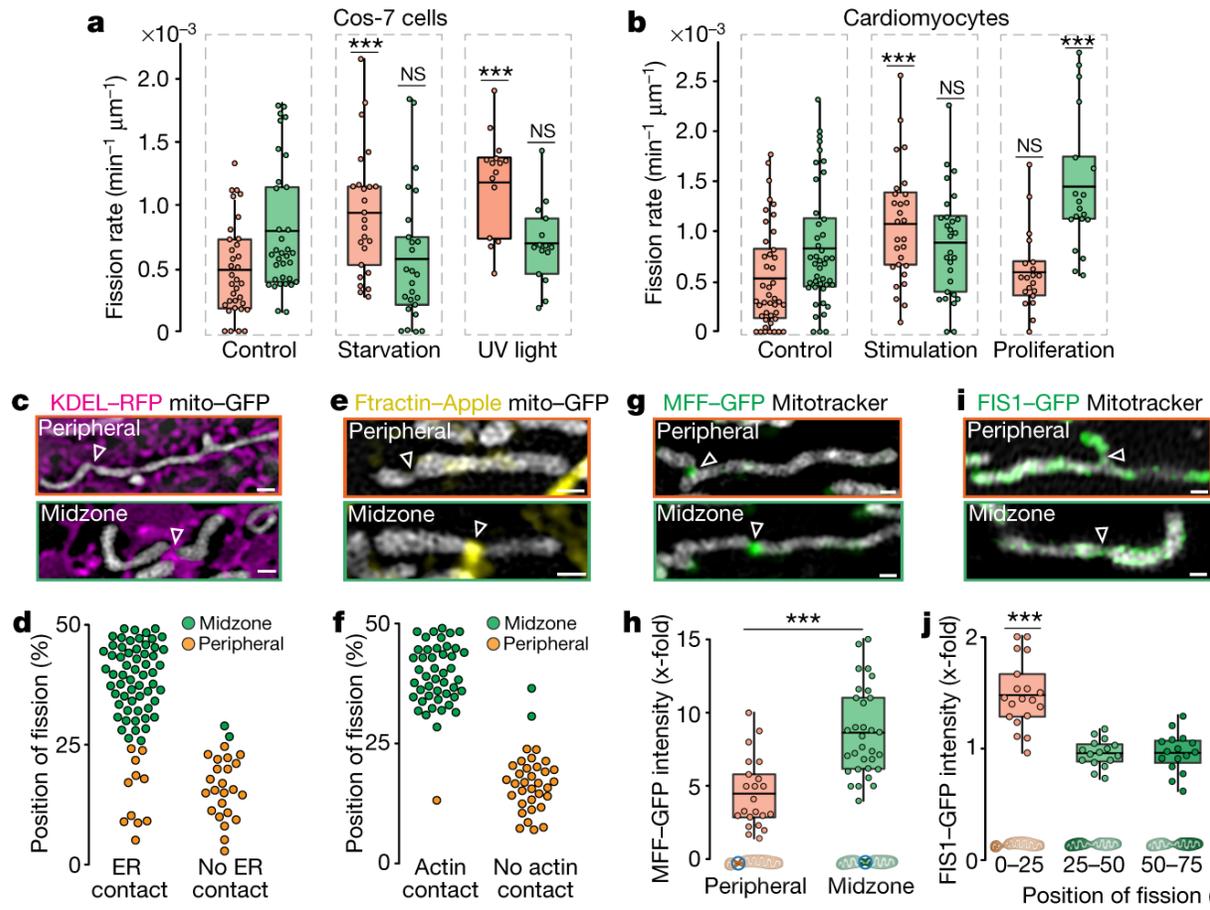
**g**, Mitochondria (mito-RFP, greyscale) and lysosomes (LAMP1-mEGFP, green) before and after peripheral or midzone fission. **h**, Position of fission in divisions contacting lysosomes or not before fission ( $n = 104$  fissions). **i**, Mitochondria (mito-RFP, greyscale) and pre-mitophagic marker (YFP-Parkin, red) before and after fission. **j**, Pre-fission YFP-Parkin fold-change in intensity as a function of fission position ( $n = 34$ ). **k**, Schema depicting the fates of daughter mitochondria from peripheral and midzone fissions. Mitochondria tracked for >100 s after fission were included. Numbers in parentheses are total numbers of events.

# Changes in mitochondrial physiology and recruitment of the fission and autophagic machinery



**Fig. Time course of physiological changes and recruitment of fission regulators** **a, b**, Time course of fluorescent signals in four examples of Cos-7 mitochondria displaying normalized TMRE intensity (**a**) and mito-R-GECO1 intensity (**b**) with corresponding SIM images in the mitochondrial compartment giving rise to the smaller daughter mitochondria before a peripheral division. **c**, Average inner membrane diameter at the constriction site at several time points before fission, measured in mito-R-GECO1 transfected Cos-7 cells during the time window where  $\text{Ca}^{2+}$  is elevated (green box in **b**,  $n = 10$  fission events). **d, e**, Time course of lysosome co-localization (**d**) and autophagosome co-localization (**e**) at constriction sites for peripheral fissions, by measuring LAMP1-mEGFP and EGFP-LC3 intensity, respectively. For EGFP-LC3 measurements, cells were pre-treated with  $10 \mu\text{M}$  CCCP to increase LC3 signals. **f**, Normalized DRP1 intensity at the constriction sites before peripheral fission in four examples of Cos-7 mitochondria with corresponding SIM images. Blue dotted lines ( $t = 0$  s) mark the time point of fission.

# Cell context-dependent modulation



与氧化损伤相关的细胞应激和高能量需求增加

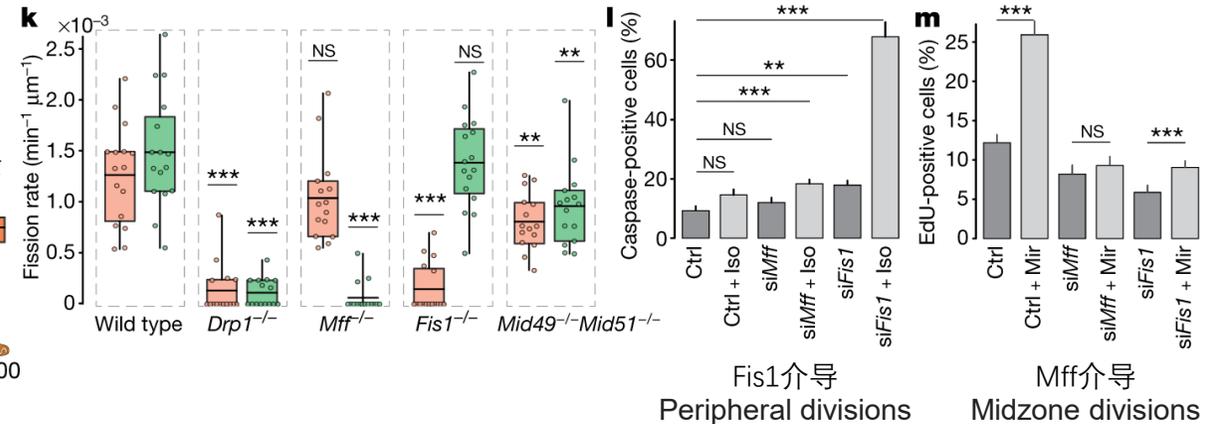
需要新线粒体生物合成的细胞增殖

Peripheral divisions的速率增加

Midzone divisions的速率增加

为了量化Mff、Fis1和MiD49/MiD51的不同作用

研究adaptor缺失的意义



**Fig. 3: Midzone and peripheral fissions are independently regulated by distinct molecular machineries.** **a**, Peripheral (orange) and midzone (green) fission rates in control, starved or UV-exposed Cos-7 cells ( $n \geq 15$  fields of view (FOV) per group). **b**, Fission rates in postnatal mouse cardiomyocytes in control, stimulated or proliferation-induced cells ( $n \geq 19$  FOV per group). **c**, SIM of mitochondria (mito-GFP, grey) and endoplasmic reticulum (ER) (KDEL-RFP, magenta) before a peripheral or midzone fission. **d**, Position of fission in mitochondria contacting the ER (left) or not (right) before fission ( $n = 93$  fissions) in Cos-7 cells. **e**, SIM of mitochondria (mito-GFP, grey) and actin (Ftractin-Apple, yellow) before peripheral and midzone fissions. **f**, Fission position in mitochondria that accumulate actin (left) or not (right) before fission ( $n = 80$  fissions). **g**, SIM of mitochondria (Mitotracker Red, grey) in U2OS cells endogenously expressing MFF-GFP for a peripheral and a midzone fission. **h**, MFF-GFP fold-change in intensity at the fission site of peripheral and midzone divisions ( $n = 54$  fissions). **i**, SIM of mitochondria (Mitotracker Red, grey) in U2OS cells endogenously expressing FIS1-GFP (green) for a peripheral and a midzone fission. **j**, FIS1-GFP fold-change in intensity over the surface of a daughter mitochondrion before fission ( $n = 35$  fissions). **k**, Peripheral (orange) and midzone (green) fission rates in wild-type, *Drp1*<sup>-/-</sup>, *Mff*<sup>-/-</sup>, *Fis1*<sup>-/-</sup> and *Mid49*<sup>-/-</sup>*Mid51*<sup>-/-</sup> double-knockout mouse embryonic fibroblasts, stained with Mitotracker Green ( $n \geq 15$  cells per group). **l**, Percentage of apoptotic (caspase 3/caspase 7-positive) cardiomyocytes ( $\pm$  s.e.m.) in control, FIS1- and MFF-depleted cells with or without stimulation with isoproterenol ( $n > 29$  FOV per group). **m**, Percentage of proliferating (Edu-positive) cardiomyocytes ( $\pm$  s.e.m.) in control, FIS1- and MFF-depleted cells with or without treatment with miR-199 (Mir) ( $n \geq 49$  FOV per group).