# Literature Report

Reporter: Liu Weiwei

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## **EDGE ARTICLE**

### Efficient and selective DNA modification on bacterial membranes

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**Dr. Mingxu You**Department of Chemistry,
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- B.S. degree in Chemistry from Peking University in 2008
- Ph.D. degree in Chemistry from the University of Florida in 2012, developing DNA-based devices for cancer diagnosis, targeted drug delivery, and cell membrane biophysical studies.
- Prior to join UMass, he pursued his postdoctoral research with Prof. Samie R. Jaffrey at Weill Medical College of Cornell University, interested in developing RNA-based fluorescent sensors for imaging metabolites and signaling molecules in live cells

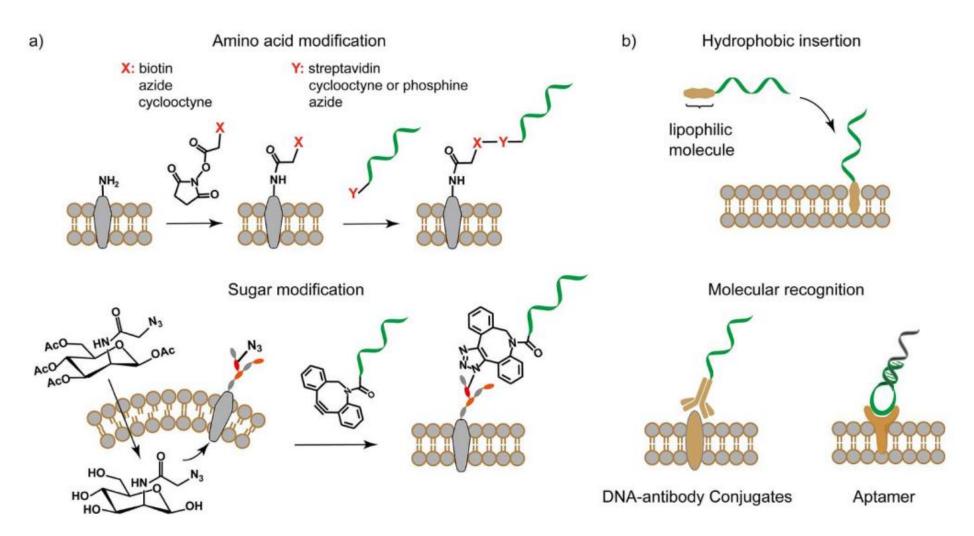
- (1) Biosensors and Bioimaging
- (2) Cell Membrane Biophysics
- (3) DNA/RNA Nanotechnology
- (4) SELEX
- (5) Photo-controlled Devices



## Introduction



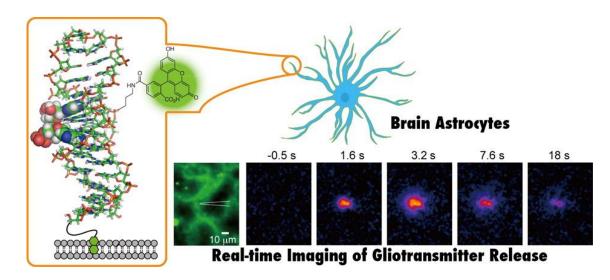
### Synthetic DNA for Cell Surface Engineering



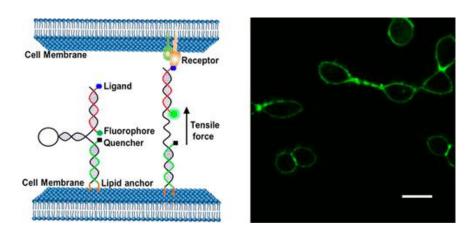


## Introduction

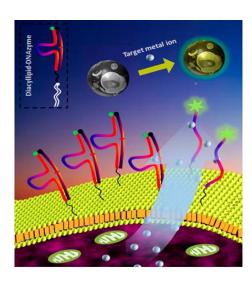
#### Lipid-DNA conjugates for cell membrane analysis



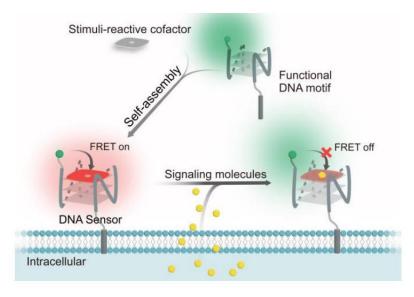
J. Am. Chem. Soc. 2012, 134, 23, 9561-9564



J. Am. Chem. Soc. 2017, 139, 50, 18182



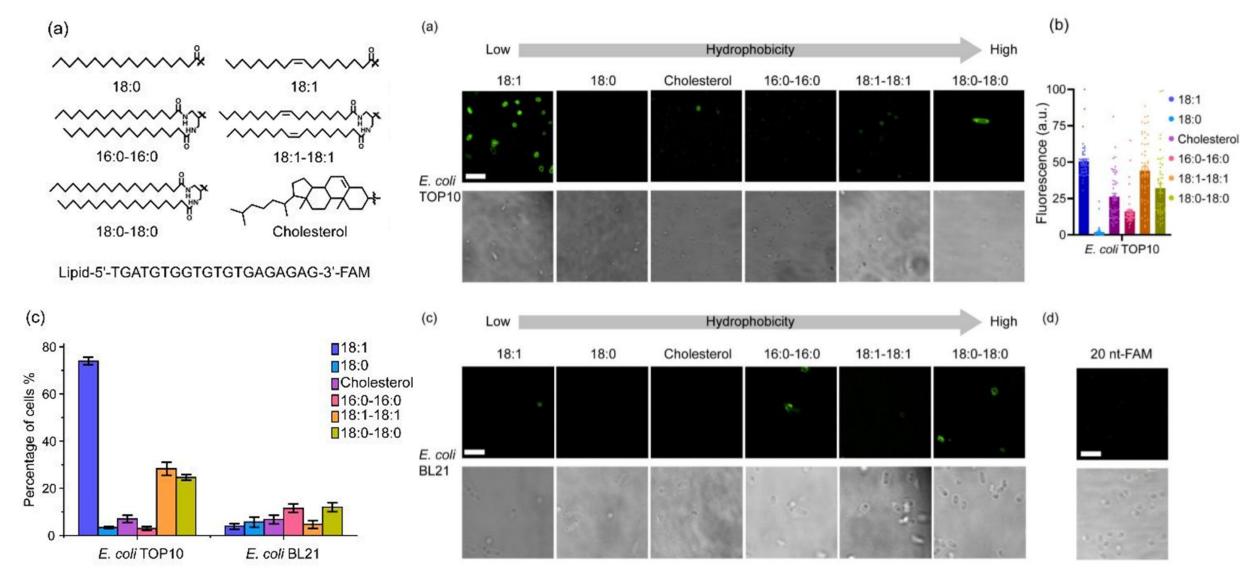
J. Am. Chem. Soc. 2014, 136, 13090-13093



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## Design and bacterial membrane insertion of lipid-DNA conjugates

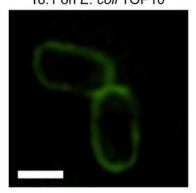




## Design and bacterial membrane insertion of lipid-DNA conjugates

#### SIM成像证明标记在外膜

(a) 18:1 on *E. coli* TOP10



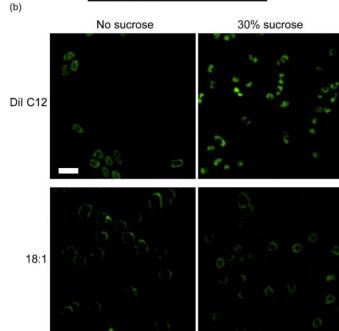
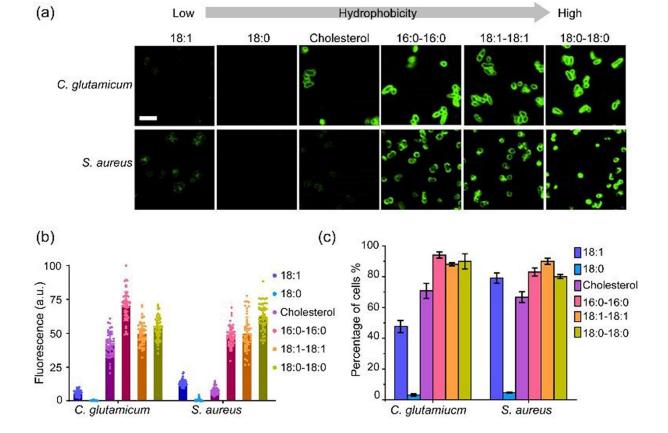


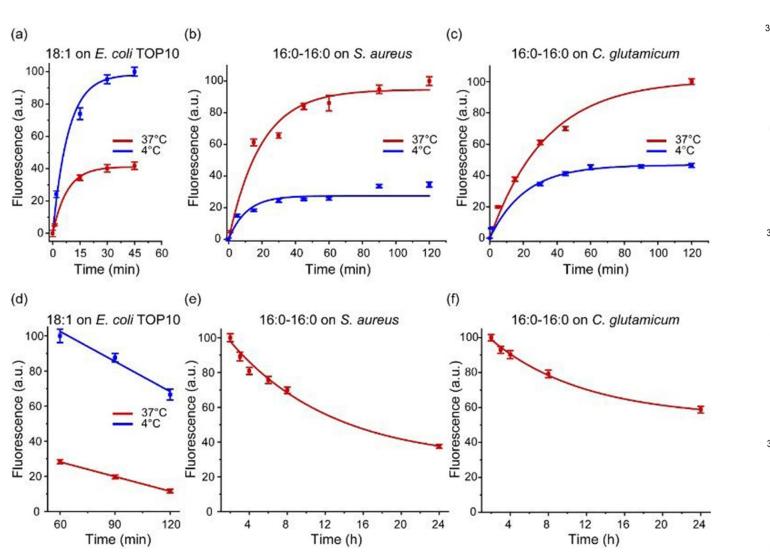
Table S2. Bacterial membrane modification percentageof each lipid-DNA conjugate.

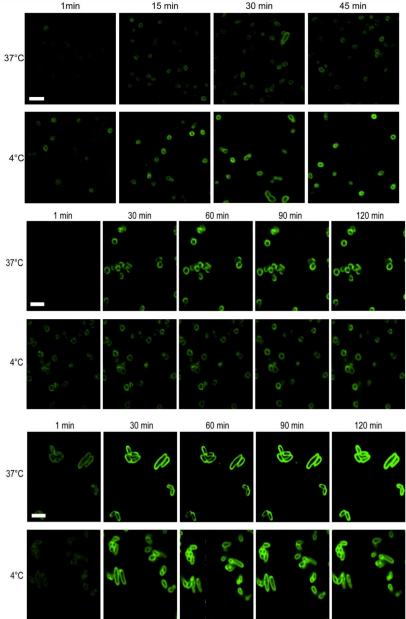
	E. coli TOP10	E. coli BL21	P. aeruginosa	C. glutamicum	S. aureus	M. luteus
18:1	84 ± 4.4	8.1 ± 0.9	<1.0	52 ± 5.5	87 ± 2.8	5.4 ± 0.5
18:0	3.9 ± 0.6	7.0 ± 1.4	<1.0	4.7 ± 0.8	5.5 ± 1.7	<1.0
Cholesterol	9.9 ± 0.9	10 ± 2.3	<1.0	71 ± 1.1	77 ± 3.7	11 ± 1.8
16:0-16:0	4.8 ± 0.4	13 ± 1.3	<1.0	94 ± 2.9	83 ± 2.1	<1.0
18:1-18:1	31 ± 1.9	6.7 ± 1.2	<1.0	88 ± 2.0	90 ± 1.2	8.8 ± 2.8
18:0-18:0	33 ± 3.2	17 ± 1.6	<1.0	90 ± 1.4	80 ± 5.2	<1.0





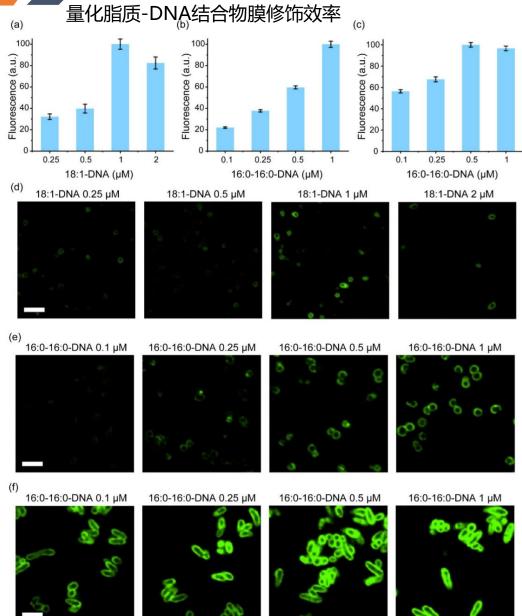
## Membrane insertion kinetics & persistence of lipid-DNA conjugates

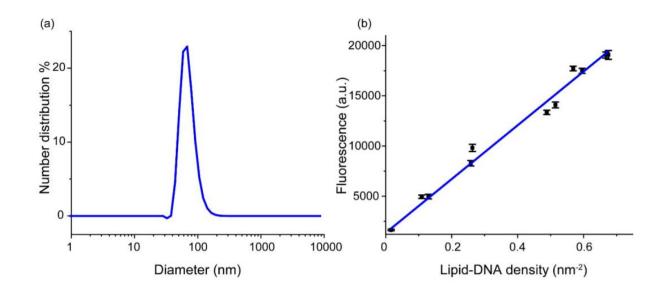






## Membrane modification efficiency of lipid-DNA conjugates



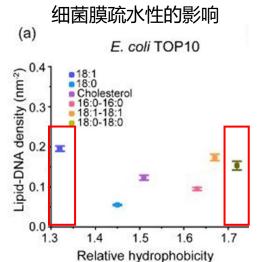


**Table S3.** Maximum bacterial membrane density of each lipid-DNA conjugate (unit: nm-2).

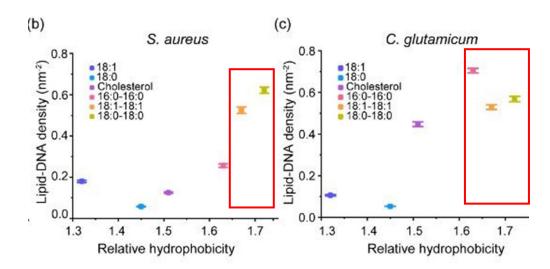
	18:1	18:0	Cholesterol	16:0-16:0	18:1-18:1	18:0-18:0
E. coli TOP10	$0.20 \pm 0.01$	$0.06 \pm 0.01$	0.12 ± 0.01	$0.10 \pm 0.01$	$0.17 \pm 0.01$	0.15 ± 0.01
S. aureus	0.18 ± 0.01	0.06 ± 0.01	0.13 ± 0.01	0.26 ± 0.01	0.53 ± 0.02	0.62 ± 0.02
C. glutamicum	0.11 ± 0.01	0.05 ± 0.01	0.45 ± 0.01	0.71 ± 0.01	0.53 ± 0.10	0.57 ± 0.01



## Effect of lipid-DNA hydrophobicity on the membrane insertion

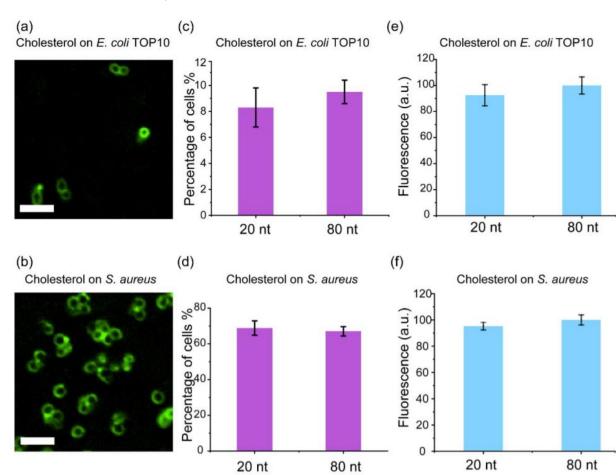


由于外膜上存在脂多糖链而具有更强的亲水性



膜含有大量高度疏水的支链氨基酸和脂肪酸

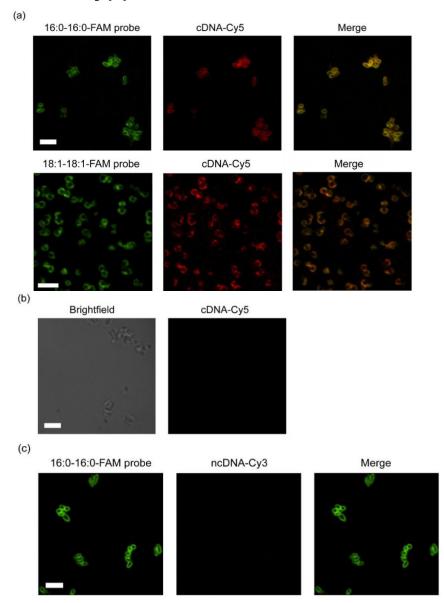
#### DNA长度的影响



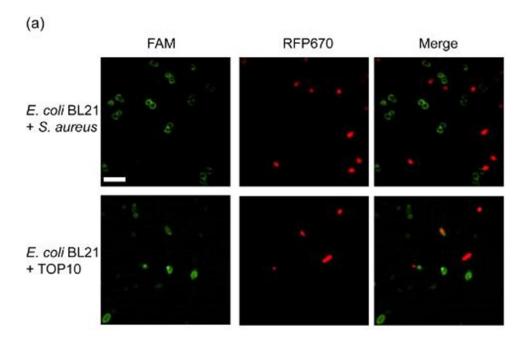


## Selective targeting and detection of bacteria

#### DNA杂交



#### 检测混合菌液中的细菌

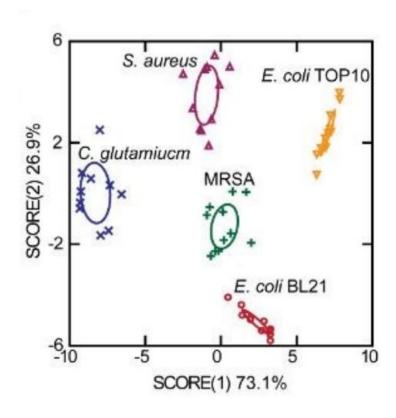


**Figure 5a.** (Top) FAM-labeled 18:1-18:1-DNA conjugate can be used to distinguish S. aureus cells from a mixture with RFP670-expressing E. coli BL21 cells. (Bottom) Similarly, FAM-labeled 18:1-DNA conjugate was used to distinguish E. coli TOP10 cells from a mixture with BL21. Here, 1  $\mu$ M of the lipid-DNA conjugate was incubated with the cell mixture for 1 h at 37°C. Scale bar, 5  $\mu$ m.

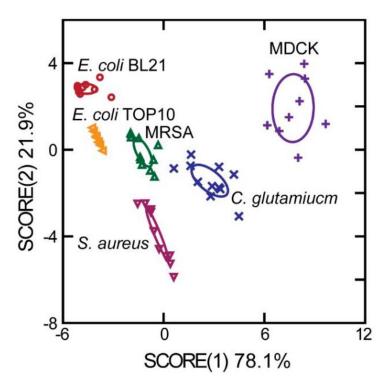


## Selective targeting and detection of bacteria

#### 检测混合菌液中的细菌



**Figure 5b**: Linear discriminant analysis based on the fluorescence response pattern of the 18:1-DNA and 16:0-16:0-DNA conjugates on five types of bacterial strains. The transformed canonical scores were plotted with 95% confidence ellipses around the centroid of each group.



**Figure S11**. Linear discriminant analysis based on the fluorescence response pattern of the 18:1-18:1-DNA and cholesterol-DNA conjugates on five types of bacterial strains and MDCK cells. The transformed canonical scores were plotted with 95% confidence ellipses and 0.001 tolerance around the centroid of each group.